

### Summary chapter 2

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#### Introduction

Although ultracentrifugation is still widely used, it is not anymore considered the gold standard methodology for Extracellular Vesicles isolation, since it does not isolate EVs efficiently, tends to alter the vesicle shape and functionality, requires expensive equipment and is time-consuming. In more than ten years of experience, we have developed and optimized methods and tools for a fast, scalable and reproducible EV purification, for addressing the EV heterogenity and high throughput solutions for biomarker discovery.

### **Ultrafilters**

For EV size separation and medium concentration



# Size Exclusion Chromatography Columns

For EV purification and removal of contaminants



### Immunoaffinity isolation

For biomarker screening and enrichment of EV subpopulations





### Chemical isolation

For a fast and simple EV precipitation from small volume of fluids







#### Tangential Flow Filters for EV concentration and purification

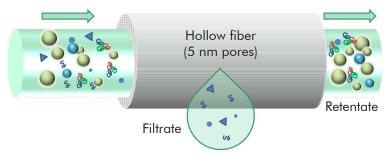
Tangential flow filtration (TFF) is a rapid and efficient method, usually used for separation and purification of biomolecules in industrial applications and can be successfully applied to isolate and separate extracellular vesicles. We provide three different typologies of filters suitable respectively for EV purification, EV concentration and Ev size-based separation.

### TFF-Easy: Filter for EV concentration and dialysis



TFF-Easy is a filter cartridge in hollow fibers made of polysulfone, which allows the particle concentration and the removal of small proteins and molecules from diluted matrices (cell conditioned media, urine, etc..), prior to the EV purification.

#### FEED FLOW



The small dimensions of the device allow to concentrate samples from 5 ml up to bigger volumes, surmounting the limit of the TFF technique which is usable for processing big volumes of fluids.

Cat. Code Filter Volume		Quantity	
TFF-Easy: EV concentration and dialysis			
HBM-TFF/1 2 ml 1 filter			
HBM-TFF/5	2 ml	5 filters	

#### References

Chetty, V. K., Ghanam, J., Anchan, S., Reinhardt, K., Brenzel, A., Gelléri, M., ... & Thakur, B. K. (2022).

Pansani, T. N. N., Phan, T. H. H., Lei, Q., Kondyurin, A., Kalionis, B., & Chrzanowski, W. (2021).

Woith, E., Guerriero, G., Hausman, J. F., Renaut, J., Leclercq, C. C., Weise, C., ... & Melzig, M. F. (2021).

Phan, T. H., Divakarla, S. K., Yeo, J. H., Lei, Q., Tharkar, P., Pansani, T. N., ... & Chrzanowski, W. (2021).

#### Characteristics

- TFF-EVs, pore zise: 50 nm.
- TFF-Easy pore size: 5 nm.
- TFF-MV pore size: 200 nm.
- Suitable for manual or mechanical use.

#### **Applications**

- Concentration of diluted fluid as cell media or urine prior to EV isolation.
- Easy removal of small molecules and ions from the EV preparation.
- EV dialysis and buffer exchange.
- High efficiency of EV isolation if coupled with SEC columns.

- Washable.
- Reusable multiple times.
- Easy to use.
- Fast concentration of EV containing matrices.



#### **Applications**

- Separation, concentration and recovery of large EVs (>150 nm).
- Large EV isolation from cell media, biofluids, plant extracts.
- Dialysis and desalting of large FVs.
- Suitable for large EV isolation from 5 ml of fluid.

#### Advantages

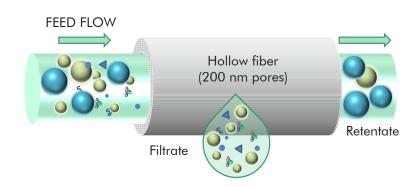
- Washable.
- Reusable multiple times.
- Sterile
- Fast separation of EVs bigger than 150 nm.

#### TFF-MV: Filter for large EV separation and purification

TFF-MV is a filter able to separate large microvesicles (MVs) by size, avoiding the separation by centrifugation at 10000g, which often causes the loss of part of small EVs. TFF-MV retains vesicles larger than 150-200 nm, whereas small EVs and circulating molecules pass in the permeate. Retained MVs can be recovered with a syringe in PBS buffer, without additional purification steps.



TFF-MV is a filter cartridge made of hollow fibers with pores of 200 nm size. It can be used manually with syringes and allows the separation of large microvesicles from small EVs (< 150 nm). It works from a mimimal amount of 5 ml of fluids up to liters of fluids.



Cat. Code Filter Volume Quantity		Quantity
TFF-MV: Separation concentration and purification of large EVs		
HBM-TFF-MV	2 ml	1 filter
HBM-TFF-MV/5	2 ml	5 filters

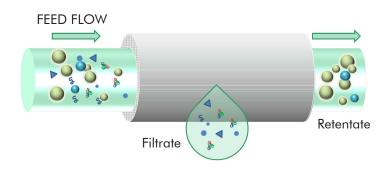




#### TFF-EVs: Fast EV purification

TFF-EVs is our next-generation filter which allows a rapid, reproducible and scalable purification of EVs, can be used on the lab bench for purifying small amount of samples (min 5 ml) or connected with a mechanical system for purifying larger volumes.

TFF-EVs is a filter cartridge made of polyethersulfonehollow fibers with pores of 50 nm size (cut off 300 kDa). The filter allows the purification of EVs and particles > 50 nm.



Avaiable in 2 sizes, the TFF-EVs Small is suitable for manual use on lab bench or under safety cabinet and allows EV purification from conditioned media or biofluids in few minutes.

The TFF-EVs Large is adapt for large scale purification and is suitable for mechanical use by peristaltic pump.



Cat. Code	Filter Volume	Quantity	
TFF-EVs: Filter for EV purification			
HBM-TFF-EVs-S	2 ml	1 filter	
HBM-TFF-EVs-L	30 ml	1 filter	

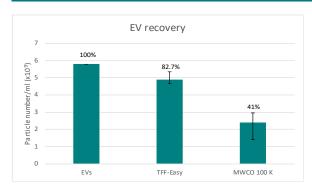
#### **Applications**

- Separation, purification and concentration of EVs from conditioned media, biofluids, plant extracts.
- Buffer exchange and removal of unbound dye.
- Depletion of FBS from bovine EVs.

- Washable.
- Reusable multiple times.
- Sterile



#### TFF-Easy: Concentration of diluted fluids with minimal loss of EVs



1. CD81 expression in concentrated vs not concentrated CCM.

30 ug of purified EVs have been diluted in 50 ml of PBS 1x and then concentrated up to 2 ml by TFF-Easy and MWCO concentrators 100 K (Millipore). The particle concentration in the final volume has been detected by NTA (Zetaview, Particle Metrix), and compared to 30 ug of EVs diluted in 2 ml of PBS 1X. TFF-Easy allowed a recovery of approximately the 83% of the particles in solution.

#### TFF-Easy allows to change the EV buffer without dialysis process

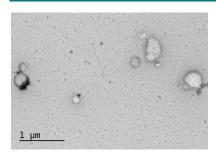
Dialysis progress	Conductivity (µS/cm)	Particle concentration (particle number/ml)
EVs in buffer 1 (PBS 1X) 5 ml	15000	5.8x10 <sup>11</sup>
1- Removal of buffer 1 by TFF	15000	
2- Injection of buffer 2 in TFF		
3- Removal of buffer 2 and buffer 1 residues	4100	
4- Injection of buffer 2		
5- Removal of buffer 2	624	
6- Injection of buffer 2		
7- Concentration of buffer 2 up to 5 ml	621	4.9x10 <sup>11</sup>

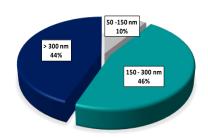
2. Process for EV dialysis with TFF-Easy.

TFF-Easy allows to dialyze EV preparation. In the process described in figure 2 we performed the EV dialysis from buffer 1 (PBS1x) to buffer 2 (NaCl 100 mM).

The TFF-Easy allows the complete removal of buffer 1, without affecting the EV concentration.

#### TFF-MV: Large EVs isolation, with minimal loss of small EVs





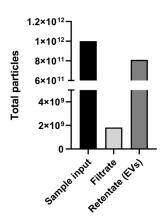
3. EM image and particle size distribution of EVs isolated with TFF-MV  $\,$ 

Currently, large MV are isolated or removed from small EVs by centrifugation (10000 g for 30 minutes), which also causes a massive loss of small vesicles. Moreover, different equipment (centrifuges, rotor angle, etc.) has impact on the final results. TFF-MV allows the removal of MV, provides their concentration and purification in one single step, skiping the centrifugation. The isolated MVs are pure and suitable for multiple analyses.



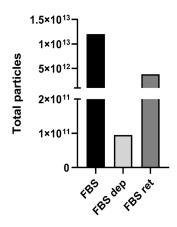


#### TFF-EVs: fast and scalable system for EV purification from CCM, biofluid, plant extracts



1x10<sup>12</sup> particles of purified EVs (HBM-PEU-100) were diluted in 20 ml of PBS 1x and then injected into TFF-EVs Small. Retentate containing EVs was recovered in 5 ml of PBS 1x. The particle content of the filtrate and retentate were analyzed by NTA (Zetaview, Particle Metrix).

#### TFF-EVs: fast and scalable system for EV purification from CCM, biofluid, plant extracts



TFF-EVs was used to depleat the FBS from EVs of bovine origine. 50 ml of raw FBS were filtered through TFF-EVs, the filtrate contained the deplated FBS, whereas bovine EVs were recovered from the retentate in 10 ml PBS 1x buffer. All the three fractions were analyzed by NTA ((Zetaview, Particle Metrix). EV depleted FBS contains only the 1% of the total particles detected in the raw FBS.

#### References

Chetty, V. K., Ghanam, J., Anchan, S., Reinhardt, K., Brenzel, A., Gelléri, M., ... & Thakur, B. K. (2022). Efficient Small Extracellular Vesicles (EV) Isolation Method and Evaluation of EV-Associated DNA Role in Cell–Cell Communication in Cancer. Cancers, 14(9), 2068.

Pansani, T. N. N., Phan, T. H. H., Lei, Q., Kondyurin, A., Kalionis, B., & Chrzanowski, W. (2021). Extracellular-Vesicle-Based Coatings Enhance Bioactivity of Titanium Implants—SurfEV. Nanomaterials, 11(6), 1445.

Woith, E., Guerriero, G., Hausman, J. F., Renaut, J., Leclercq, C. C., Weise, C., ... & Melzig, M. F. (2021). Plant extracellular vesicles and nanovesicles: Focus on secondary metabolites, proteins and lipids with perspectives on their potential and sources. International journal of molecular sciences, 22(7), 3719.

Phan, T. H., Divakarla, S. K., Yeo, J. H., Lei, Q., Tharkar, P., Pansani, T. N., ... & Chrzanowski, W. (2021). New Multiscale Characterization Methodology for Effective Determination of Isolation–Structure–Function Relationship of Extracellular Vesicles. Frontiers in Bioengineering and Biotechnology, 9, 358.

Useckaite, Z., Mukhopadhya, A., Moran, B., & O'Driscoll, L. (2020). Extracellular vesicles report on the MET status of their cells of origin regardless of the method used for their isolation. Scientific reports, 10(1), 1-11.

Mangino, G., Iuliano, M., Carlomagno, S., Bernardini, N., Rosa, P., Chiantore, M. V., ... & Romeo, G. (2019). Interleukin 17A affects extracellular vesicles release and cargo in human keratinocytes. Experimental dermatology, 28(9), 1066-1073.



#### Characteristics

- Filter membrane in polyethersulfone.
- MWCO: 100 kDa.
- Concentration volume: from 2.5 ml to 0.050 ml.
- Reverse ultrafiltration (opposite direction to the centrifugal force).

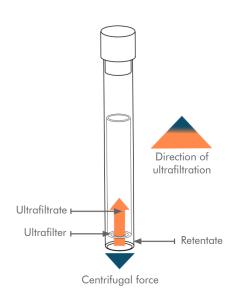
#### **Applications**

- Concentration of samples containing EVs or nanopartilces.
- Concentration of small volumes of diluted fluids.
- EV dialysis for changing buffer conditions.
- Removal of contaminants and small molecules (unbound dyes).

#### EV-Spinner: ultrafiltration concentrator

EV-Spinner is a 2.5 ml non-stick ultrafiltration (UF) concentrator to enhance the recovery of extracellular vesicle (EVs) during the concentration or buffer exchange step. The ultrafiltration works in the opposite direction to the centrifugal force, providing higher particle recovery. The low protein binding membrane (polyether-sulphone) reduces the EV loss, compared to V-shape concetrators, and the reverse design of the EV-spinner ensures that the filter does not clog.





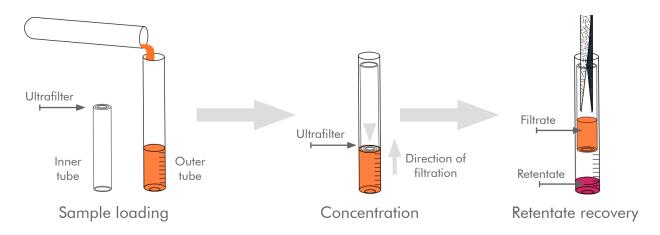
- Non-stick.
- Easy recovery of concentrated material.
- Suitable for multiple washing.

Cat. Code Description		
EV-Spinner ultrafiltration concentrator		
HBM-EVS-24	EV-Spinner 100 kDa MWCO concentrators, 24 pieces	
HBM-EVS-48	EV-Spinner 100 kDa MWCO concentrators, 48 pieces	

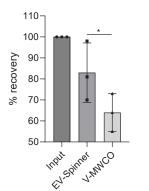




#### Reverse ultrafiltration for maximum recovery of extracellular vesicles and nanoparticles

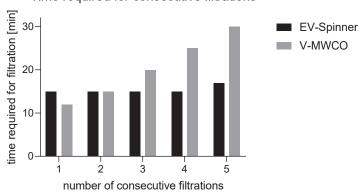


#### Percent of recovered EVs



Up to 98% of EV recovery was observed with 2.5 ml of EV solution concentrated up to 0.25 ml. Symbols are biological repeats, bars indicate means and error-bars are SDs.

#### Time required for consecutive filtrations



EV-Spinner allows consecutive concentrations (from 2.5~ml up to 0.25~ml) with minimun clogging of the filter. The clogging of the filter was measured by the time necessary for concetrating a solution of EVs from 2.5~ml up to 0.25~ml.



#### Characteristics

- New gel matrix for EV purity improvement.
- Purification up to 20 ml volume of fluid.

#### PURE-EVs Size Exclusion Chromatography Columns

Size Exclusion Chromatography (SEC) is an efficient method for isolating and purifying Extracellular Vesicles (EVs) from different fluids, not affecting the original shape and functionality of the vesicles. We have developed a set of SEC columns which allow the EV purification from small (100  $\mu$ l) and large volumes (up to 20 ml) of fluids. The EV purification process with PURE-EV columns is very fast, taking approximately 15 minutes of time.

#### **Applications**

- Extracellular vesicles isolation from cell media, biofluids and plant extracts.
- Purification of EVs from contaminants.
- Dye excess removal post EV labeling process.

Cat. Code	Volume	Columns		
PUR	E-EV: Size Exclusion Ch	nromatography columns		
HBM-PEV-5	500 μl - 2 ml	5 Columns		
HBM-PEV-10	500 μl - 2 ml	10 Columns		
miniPl	miniPURE-EV: Size Exclusion Chromatography columns			
HBM-mPEV-10	100 μl - 500 μl	10 Columns		
HBM-mPEV-20	100 μl - 500 μl	20 Columns		
maxiPURE-EV: Size Exclusion Chromatography columns				
HBM-mxPEV-3	1 ml - 20 ml	3 Columns		
HBM-mxPEV-6	1 ml - 20 ml	6 Columns		

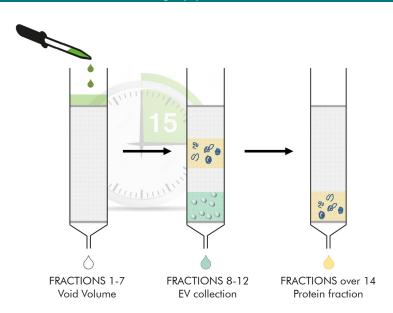
- Easy and fast protocol (turnaround time approximately 15 minutes).
- Isolate EVs from small sample volumes.
- Reusable up to 5 times.
- Long term stability at 4°C.



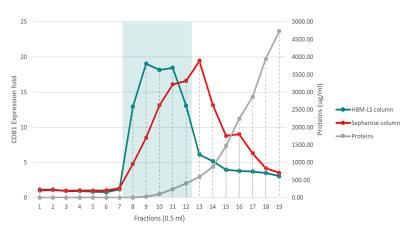




#### PURE-EVs: isolation of highly pure extracellular vesicles in approximately 15 minutes

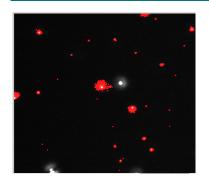


PURE-EVs column was rinsed with 1 ml of cell conditioned media from HCT116 cells, preconcentrated with TFF-Easy 10 fold. 24 fractions (500  $\mu$ l each one) have been collected and analyzed by ELISA ExoTEST<sup>TM</sup> assay (see section 3, ExoTEST quantification kit) and by BCA test for determining EVs and total protein content. Results were compared with a column filled with Sepharose CL2B (GE Healthcare). EVs are eluted in fractions 8 - 12, whereas the peak corresponding to protein fraction starts from fraction 14 (Fig 4).

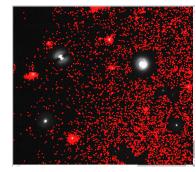


4. EV purification with PURE-EVs column (green line) vs column filled with Sepharose CL2B

#### mini-PURE-EVs: optimal method for removing the dye excess post EV labeling



5. Dye excess removed by mini-PURE-EVs



6. Dye excess not removed

 $10\,\mu g$  of purified EVs from HCT116 cells were labeled by the membrane dye Cell Mask Green. The excess of the dye has been removed from the EV preparation using a mini-PURE-EVs column. The background removal has been detected by NTA with Zetaview (Particle Metrix) (Fig 5,6).



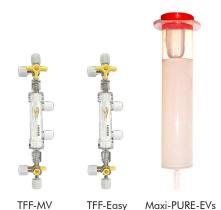
#### **Applications**

- Separation and purification of both large (> 150 nm) and small EVs.
- EV concentration and buffere exchange.
- Suitable for conditioned media and diluted biofluids (Urine).

#### PURE-EVs COMBO kits: EV purification from diluted fluids

PURE-EVs PLUS and PURE-EVs COMPLETE are kits which combine the ability of the TFF filters to concentrate diluted fluids to the capacity of the PURE-EVs columns to purify EVs from circulating proteins. The Combo kits are the perfect solution for people who isolate EVs from fluids as cell conditioned media or urine and want to obtain a high recovery and separation of small and large vesicles.

PURE-EVs COMPLETE: Double TFF and SEC for scalable and reproducible EV isolation and size fractionation from diluted fluids.



PURE-EVs PLUS: TFF-Easy and SEC combination for EV isolation and size fractionation.



TFF-Easy Maxi-PURE-EVs

Cat. Code	Volume	Columns
PURE-EV COMPLETE: Complete EV purification and fractionation		
HBM-PEV-5	500 μl - 2 ml	5 Columns
HBM-PEV-10	500 μl - 2 ml	10 Columns
PURE-EV PLUS:		
HBM-mPEV-10	100 μl - 500 μl	10 Columns
HBM-mPEV-20	100 μl - 500 μl	20 Columns





#### References

Yim, K. H. W., Borgoni, S., & Chahwan, R. (2022). Serum extracellular vesicles profiling is associated with COVID[19 progression and immune responses. Journal of Extracellular Biology, 1(4), e37.

Leal, C. L. V., Cañón-Beltrán, K., Cajas, Y. N., Hamdi, M., Yaryes, A., de la Blanca, M. G. M., ... & Rizos, D. (2022). Extracellular vesicles from oviductal and uterine fluids supplementation in sequential in vitro culture improves bovine embryo quality.

Sun, C. Y., Chen, G. D., He, B. C., Fu, W. E., Lee, C. H., Leu, Y. W., & Hsiao, S. H. (2022). Dysregulated HIC1 and RassF1A expression in vitro alters the cell cytoskeleton and exosomal Piwi-interacting RNA. Biochemical and Biophysical Research Communications.

Rana, N., Suliman, S., Al-Sharabi, N., & Mustafa, K. (2022). Extracellular Vesicles Derived from Primed Mesenchymal Stromal Cells Loaded on Biphasic Calcium Phosphate Biomaterial Exhibit Enhanced Macrophage Polarization. Cells, 11(3), 470.

Nikiforova, N., Chumachenko, M., Nazarova, I., Zabegina, L., Slyusarenko, M., Sidina, E., & Malek, A. (2021). CM-Dil Staining and SEC of Plasma as an Approach to Increase Sensitivity of Extracellular Nanovesicles Quantification by Bead-Assisted Flow Cytometry. Membranes, 11(7), 526.

Cañón-Beltrán, K., Hamdi, M., Mazzarella, R., Cajas, Y. N., Leal, C. L., Gutiérrez-Adán, A., ... & Rizos, D. (2021). Isolation, Characterization, and MicroRNA Analysis of Extracellular Vesicles from Bovine Oviduct and Uterine Fluids. Next Generation Culture Platforms for Reliable In Vitro Models: Methods and Protocols. 219-238.

Han, P., Bartold, P. M., & Ivanovski, S. (2021). The emerging role of small extracellular vesicles in saliva and gingival crevicular fluid as diagnostics for periodontitis. Journal of Periodontal Research.

Han, P., Bartold, P. M., Salomon, C., & Ivanovski, S. (2021). Salivary Outer Membrane Vesicles and DNA Methylation of Small Extracellular Vesicles as Biomarkers for Periodontal Status: A Pilot Study. International Journal of Molecular Sciences, 22(5), 2423.

Alameldin, S., Costina, V., Abdel-Baset, H. A., Nitschke, K., Nuhn, P., Neumaier, M., & Hedtke, M. (2021). Coupling size exclusion chromatography to ultracentrifugation improves detection of exosomal proteins from human plasma by LC-MS. Practical Laboratory Medicine, e00241.

Hamdi, M., Mazzarella, R., Cañon-Beltrán, K., Cajas, Y. N., Leal, C. L. V., Gutiérrez-Adán, A., ... & Rizos, D. (2021). 36 Analysis of miRNA content of oviduct and uterine extracellular vesicles across the bovine estrous cycle. Reproduction, Fertility and Development, 33(2), 125-125.

Yaryes Estaque, A. A. (2020). Competence of oviductal and uterine Extracellular Vesicles in sequential culture of in vitro bovine embryos

Han, P., Bartold, P. M., Salomon, C., & Ivanovski, S. (2020). Salivary Small Extracellular Vesicles Associated miRNAs in Periodontal Status—A Pilot Study. International Journal of Molecular Sciences, 21(8), 2809.

Leal, C., Cañon-Beltrán, K., Cajas, Y., Gallego, P., Beltrán-Breña, P., Hamdi, M., ... & Rizos, D. (2020). 76 Extracellular vesicles from oviduct and uterus in sequential culture improve the quality of bovine embryos produced in vitro. Reproduction, Fertility and Development, 32(2), 164-164.

Yang, D., Zhang, W., Zhang, H., Zhang, F., Chen, L., Ma, L., ... & Tran, P. H. (2020). Progress, opportunity, and perspective on exosome isolation-efforts for efficient exosome-based theranostics. Theranostics, 10(8), 3684.



#### Immunoaffinity isolation of Extracellular Vesicles

Addressing the EV heterogenity is becoming an important issue, in particular for the different roles that Extracellular Vesicles have in pathological processes as cancer, infection, or neurodegenerative diseases.

HBM-LS provides pre-coated ELISA Immunoplates and Latex or Magnetic Immunobeads for the capture and enrichment of total or specific EV-subpopulations.

#### **Applications**

- Multiple profiling of EV markers from a single sample or screening of a large number of samples.
- EV capture and quantification from human biofluids (plasma, serum, urine, saliva).
- Suitable for nucleic acid extraction from immunocaptured EVs.

#### Immunoplates for EV capture and isolation



HBM-LS Immunoplates are 96 multiwell plates covalently pre-coated with specific EV-binding antibodies allowing the capture and isolation of vesicles from different sources (cell supernatant, human plasma, serum, urine and saliva). We developed different types of plates for capturing the total or for enriching specific EV subpopulations (tumoral, neural, glial derived). Plates are blocked and stabilized for long-term storage.

#### Advantages

- Ready to use.
- Long term storage (up to 2 years).
- No EV pre-purification required
- Small amount of sample required (100 μl per well).
- Flexibility in designing a multiplexing assay.
- Open platform for customized coating solutions.

#### Immunoplates for Total EV capture and isolation

Cat. Code	Immunoplate	Antibody	Recommended for		
ELISA Immunop	ELISA Immunoplate for CD9 positive Extracellular Vesicles capture				
HBM-POS-CC/T1	Transparent		l l	Human Plasma,	
HBM-POS-CC/W1	White	Mouse Monoclonal	Serum, Urine		
ELISA Immunop	late for CD63 po	ositive Extracellular Vesicle	es capture		
HBM-POC-CC/T1	Transparent	Anti Human CD63	Cell conditioned		
HBM-POC-CC/W1	White	Mouse Monoclonal	media, Biofluids		
	Custommad	e Immunoplate			
Plates can be covalentely coated with EV binding antibodies, choosen from our anti- body list or sent by customers. Plates are available in transparent and white format.					
Storage conditions					
Unopened: 2 years, stored at 4°C. Opened: 6 month stored at 4°C					
Material amount					
$100\mu l$ of sample per well. Whole human plasma and serum can be loaded for vesicle capture. Using urine and cell media, it is recommended to concentrate the sample 10 folds, before loading the sample on the plate.					
Packaging information					
			·		

Immunoplates are individually sealed in an opaque aluminium ziplock bag, compliant

to pharmaceutical regulations. Easy to open and reseal.





#### Immunoplates for enrichment of Tumor-derived EVs

Immunoplates coated with antibodies against TM9SF4 or EpCAM, two proteins widely expressed in tumor tissues and in tumor-derived EVs.

TM9SF4: TM9SF4 is a membrane protein involved in the activation of V-ATPases in conditions of elevated intracellular concentration of H+ as a consequence of elevated fermentation of sugars (Warburg effect). Ref: Lozupone F. et al. 2015

**EpCAM:** EpCAM is a transmembrane glycoprotein highly expressed in rapidly growing epithelial tumors. It plays an important role in localization of EVs in numerous physiological and phatological processes. Ref: Jiang L. et al. 2017; Yu L. et al. 2013

Cat. Code	Immunoplate	Antibody	Recommended for
ELISA Immunople	ate for TM9SF4 F	ositive Extracellular Vesic	les capture
HBM-PTF-CC/T1	Transparent	Anti Human TM9SF4	Human Plasma,
HBM-PTF-CC/W1	White	Mouse Monoclonal	Serum
ELISA Immunoplate for EpCAM Positive Extracellular Vesicles capture			
HBM-PTE-CC/T1	Transparent	Anti Human EpCAM Mouse Monoclonal	Human Plasma,
HBM-PTE-CC/W1	White		Serum

#### Immunoplates for enrichment of Neural and Glial EVs

Immunoplates coated with antibodies against EV surface antigens and indicative of neurological or glial origin.

L1CAM: L1CAM is a neural cell adhesion molecule, implicated in cell migration, adhesion and neuronal differentiation. L1CAM is highly expressed in EVs from neural origin and cna be used for enrichment of neural derived EV subpopulation. Ref: Mustapic et al. 2017

PLP1: PLP1is the major myelin protein from the central nervous system and plays an importan role in the formation and the maintenance of the myelin structure. EVs derived from glial cells are characterized by the presence of high levels of PLP1 protein.

Ref: Frühbeis et al. 2012

Cat. Code	Immunoplate	Antibody	Recommended for
ELISA Immunopl	ate for L1CAM P	ositive Extracellular Vesicl	es capture
HBM-PNF-CC/T1	Transparent	Anti Human L1CAM-	Human Plasma,
HBM-PNF-CC/W1	White	Mouse Monoclonal	Serum, Cell media
ELISA Immunoplate for PLP1 Positive Extracellular Vesicles capture			
HBM-PGF-CC/T1	Transparent	Anti Human PLP1 Mouse Monoclonal	Human Plasma,
HBM-PGF-CC/W1	White		Serum, Cell media

#### **Applications**

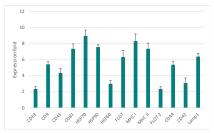
- Capture and enrichment of Tumor derived EVs subpopulations.
- Suitable for nucleic acid extraction from immunocaptured EVs.
- Profiling of cancer related biomarkers.

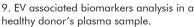
- Ready to use.
- Long term storage (up to 2 years).
- No EV pre-purification required
- Small amount of sample required (100 μl per well).
- Open platform for customized coating solutions.

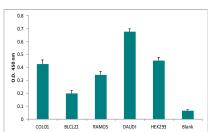


#### Immunoplates allow EV phenotyping without vesicle pre-purification steps

ELISA Immunoplates can be used for quantitative and qualitative analysis of EV-associated proteins. The plate is able to capture EVs from raw biologic material (plasma, serum, cell medium, etc.). No significant cross-reactivity is observed with soluble antigens.



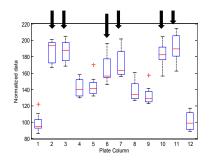


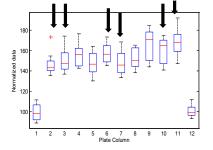


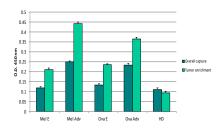
10. CD63 profiling of different cell derived EVs.

#### Enrichment of Tumor-derived Extracellular Vesicles

Immunoplates for tumor-derived EV enrichment (TM9SF4 coated) are able to distinguish cancer patients (black arrows) from healthy controls. The enrichment of tumor-derived EVs from cancer patient (Melanoma, Ovary) is detectable when the TM9SF4 coated plate is compared with a plate coated with CD9 (fig11).



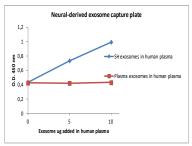




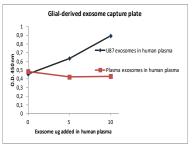
11. Enrichment in Tumor-derived EVs in early and late stage melanoma (Mel E; Mel ADV) and ovary carcinoma (Ova E, Ova ADV).

#### Enrichment of Neural or Glial derived Extracellular Vesicles

Immunocapture enrichment of neuraland glial-derived EVs purified from SK-N-SH and U87 cell lines and spiked in human plasma. Comparison was done with purified plasma EVs spiked in human plasma (fig 12 and fig 13).



12. Enrichemnt of SK-N-SH derived EVs spiked in human plasma from healthy donors using HBM-PNF



13. Enrichmentof U87 derived EVs spiked in human plasma from healthy donors using HBM-PGF





#### Immunobeads for Total EV isolation



Latex or Magnetic immunobeads are covalently coupled with antibodies against common EV surface antigens (CD9, CD63). They allow capturing EV from human biofluids (tested for plasma, serum and urine) and cell culture media without the necessity of pre-purification steps. The kit includes a Beads Elution buffer, for detaching captured EVs from antibodies and a Regeneration buffer, for regenerating the beads that can be reused ones more. Beads are sold in package of 10 reactions and are available in 2 sizes (0.4 and 1 micron diameter).

Cat. Code	Bead diameter	Antibody	Recommended for		
Immunobe	Immunobeads for CD9 Positive Extracellular Vesicles capture				
HBM-BOLF-CC/10-04	0.4 micron	Anti Human CD9	EV pheno/genotyping		
HBM-BOLF-CC/10-1	1 micron	Mouse Monoclonal	FACS analysis		
Immunobe	ads for CD63 Posi	tive Extracellular Vesicl	es capture		
HBM-BOLC-CC/10-04	0.4 micron	Anti Human CD63	EV pheno/genotyping		
HBM-BOLC-CC/10-1	1 micron	Mouse Monoclonal	FACS analysis		
Immunobeads	for Mouse Extrace	llular Vesicles capture	(CD9 Positive)		
HBM-BMLF-CC/10-04	0.4 micron	Anti Mouse CD9	EV pheno/genotyping		
HBM-BMLF-CC/10-1	1 micron	Mouse Monoclonal	FACS analysis		
Custommade Latex or Magnetic Immunobeads					
Beads can be covalentely coated with EV binding antibodies, choosen from our anti- body list or sent by customers. Magnetic or Latex beads are available.					
Storage Condition					
Store the immunobeads and buffers at 4 - 8° C.					
Conditions required					
Recommended starting volume from 0.1 ml - 0.5 ml of plasma, from 0.5 ml to 1 ml of serum.					

Concentrated (10X) urine and cell culture medium samples are recommended prior

capture according to our suggested protocol (see page 21, TFF-Easy).

#### **Applications**

- Total EV isolation from cell culture media, human or mouse biofluids (tested for plasma, serum, urine).
- Total EV isolation from mouse biofluids (tested for plasma and serum).
- Downstream marker profiling.
- Nucleic acids extraction
- EV elution from immunobeads

- Ready to use.
- Small sample volume of biofluid or cell culture medium.
- No ultracentrifugation or other methods for vesicle purification required.
- Supplied with buffer for EV elution from beads.
- Immunobeads can be regenerated with Beads Regeneration Buffer and reused.



#### **Applications**

- Capture and enrichment of human EV subpopulation (tumorderived).
- Downstream EV marker profiling.
- Nucleic acids extraction
- EV elution from immunobeads

#### Immunobeads for Tumor-derived EV capture

Latex or Magnetic immunobeads are covalently coupled with antibodies against EV surface antigens (TM9SF4 or EpCAM) associated with pathological conditions (cancer). They allow to pull down tumor-derived EV from human biofluids, thus providing a potential new platform for the research in circulating tumor biomarker.

Cat. Code	Bead diameter	Antibody	Recommended for	
Immunobeads for TM9SF4 Positive Extracellular Vesicles capture				
HBM-BTLF-CC/10-04	0.4 micron	TMOSE/ Pabbit	EV pheno/genotyping	
HBM-BTLF-CC/10-1	1 micron		FACS analysis	
Immunobeads for EpCAM Positive Extracellular Vesicles capture				
HBM-BTLE-CC/10-04	0.4 micron	Anti Human EpCAM Mouse Monoclonal	EV pheno/genotyping	
HBM-BTLE-CC/10-1	1 micron		FACS analysis	
Packaging information				
Immunobeads (10 reactions) are supplied with Exosome Elution Buffer, for eluting intact exosomes from beads and with Bead Regeneration Buffer, for regenerating immunobeads that can be reused twice more.				

#### References

Anastasi, F., Masciandaro, S. M., Carratore, R. D., Dell'Anno, M. T., Signore, G., Falleni, A., ... & Bongioanni, P. (2021). Proteomics Profiling of Neuron-Derived Small Extracellular Vesicles from Human Plasma: Enabling Single-Subject Analysis. International journal of molecular sciences, 22(6), 2951.

Zocco, D., Bernardi, S., Novelli, M., Astrua, C., Fava, P., Zarovni, N., ... & Foroni, C. (2020). Isolation of extracellular vesicles improves the detection of mutant DNA from plasma of metastatic melanoma patients. Scientific reports, 10(1), 1-12.

Luddi, A., Zarovni, N., Maltinti, E., Governini, L., De Leo, V., Cappelli, V., ... & Piomboni, P. (2019). Clues to Non-Invasive Implantation Window Monitoring: Isolation and Characterisation of Endometrial Exosomes. Cells, 8(8), 811.

Ambrosio, M. R., Vernillo, R., De Carolis, S., Carducci, A., Mundo, L., Ginori, A., ... & Cricca, M. (2019). Putative role of circulating Human Papillomavirus DNA in the development of primary squamous cell carcinoma of the middle rectum: a case report. Frontiers in oncology, 9, 93.

Storci, G., De Carolis, S., Papi, A., Bacalini, M. G., Gensous, N., Marasco, E., ... & Sarnelli, A. (2019). Genomic stability, anti-inflammatory phenotype, and up-regulation of the RNAseH2 in cells from centenarians. Cell Death & Differentiation, 1.

Trivedi, M. S., & Abreu, M. (2018). Nucleic Acid Profiling in Tumor Exosomes. In Diagnostic and Therapeutic Applications of Exosomes in Cancer (pp. 93-117).

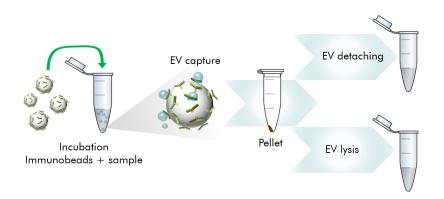
Zarovni, N., Corrado, A., Guazzi, P., Zocco, D., Lari, E., Radano, G., ... & Chiesi, A. (2015). Integrated isolation and quantitative analysis of exosome shuttled proteins and nucleic acids using immunocapture approaches.

Jia, S., Zocco, D., Samuels, M. L., Chou, M. F., Chammas, R., Skog, J., ... & Kuo, W. P. (2014). Emerging technologies in extracellular vesicle-based molecular diagnostics. Expert review of molecular diagnostics, 14(3), 307-321.

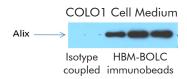




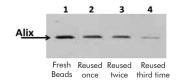
#### Immunobeads allow EVs capture and multiple downstraem analyses



Following incubation, beads can be recovered by centrifugation, resuspended in Laemmli buffer for SDS-PAGE and western blotting analysis (fig 14, 15) or in appropriate lysis buffer for nucleic acid analysis (fig 16). Alternatively, the vesicles can be eluted from the beads with the Elution Buffer and used for downstream applications such as ELISA or NTA. Eluted beads can be regenerated with Bead Regeneration Buffer and reused for capturing exosomes twice more (fig 15).

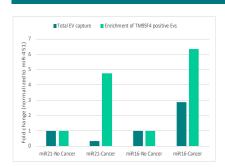


14. Alix expression by western blotting of exosomes captured on HBM-BOLC immunobeads from COLO1 cell supernatant vs isotype coupled beads.

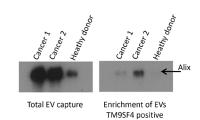


15. Western Blotting analysis of immunocaptured exosomes on beads.

#### TM9SF4 coated immunobeads enrich Tumor-derived EVs in cancer patient



16. Enrichment of miR21 and miR16 in cancer when TM9SF4 coated beads are used



17. Comparison of detection in WB of total and TM9SF4 positive EVs

Latex or Magnetic Immunobeads can be used for capturing EVs from raw biofluids, followed by RNA isolation. The enrichment of miRNA cancer associated (miR16 and miR21) is highly detectable when Immunobeads for capture of Tumor-derived EVs (HBM-BTF; TM9SF4 coated) are used.



#### **Applications**

- Single step isolation of EVs from multiple fluids
- Isolate the overall vesicles population in a sample.
- Isolated EVs are suitable for nucleic acid extraction and profiling.
- Isolated EVs are suitable for protein profiling (WB, ELISA, FACS).

#### EXO-Prep: one step EV isolation reagent



EXO-Prep is a fast and convenient method of Extracellular Vesicle isolation from biofluids, cell culture supernatants, plant extracts. Isolation with EXO-Prep is based on chemical precipitation. Samples are incubated with EXO-Prep solution on ice so that EVs will precipitate following centrifugation. The obtained pellet can be resuspended in PBS 1X or deionized water. The protocol is user-friendly, time-saving (around 1 hour), and does not require capital laboratory equipment. Isolated vesicles are in particular suitable for isolation of nucleic acid associated to EVs.

#### Advantages

- Time and money saving.
- No ultracentrifugation required.
- Easy and fast protocol.
- Isolate EVs from small volumes of sample (as low as 100 μl of plasma).
- Easy to store and ship (4°C).

Cat. Code	Volume	Reactions	
EXO-Prep for Exosome Isolation from Plasma and Serum			
HBM-EXP-B5	5 ml	180 reactions Plasma, 80 reactions Serum	
EXO-Prep for Exosome Isolation from Cell Media			
HBM-EXP-C25	25 ml	25 reactions	
EXO-Prep for Exosome Isolation from Urine			
HBM-EXP-U25	30 ml	25 reactions	

#### References

Zhang, H., Zhang, Q., Deng, Y., Chen, M., & Yang, C. (2022). Improving Isolation of Extracellular Vesicles by Utilizing Nanomaterials. Membranes, 12(1), 55.

Ogino, N., Takahashi, H., Nagaoka, K., Harada, Y., Kubo, M., Miyagawa, K., ... & Ogino, K. (2021). Possible contribution of hepatocyte secretion to the elevation of plasma exosomal arginase-1 in high-fat diet-fed mice. Life Sciences, 278, 119588.

Slyusarenko, M., Nikiforova, N., Sidina, E., Nazarova, I., Egorov, V., Garmay, Y., ... & Malek, A. (2021). Formation and Evaluation of a Two-Phase Polymer System in Human Plasma as a Method for Extracellular Nanovesicle Isolation. Polymers, 13(3), 458.

Bai, G., Matsuba, T., Niki, T., & Hattori, T. (2020). Stimulation of THP-1 Macrophages with LPS Increased the Production of Osteopontin-Encapsulating Exosome. International journal of molecular sciences, 21(22), 8490.

Lu, C. H., Chen, Y. A., Ke, C. C., Chiu, S. J., Chen, C. C., Hsieh, Y. J., ... & Liu, R. S. (2020). Preclinical Characterization and In Vivo Imaging of 111 In-Labeled Mesenchymal Stem Cell–Derived Extracellular Vesicles. Molecular imaging and biology, 1-11.

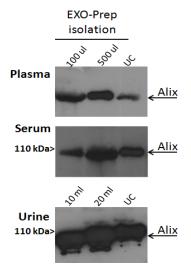
Zocco, D., Bernardi, S., Novelli, M., Astrua, C., Fava, P., Zarovni, N., ... & Foroni, C. (2020). Isolation of extracellular vesicles improves the detection of mutant DNA from plasma of metastatic melanoma patients. Scientific reports, 10(1), 1-12.

Carnino, J. M., Lee, H., & Jin, Y. (2019). Isolation and characterization of extracellular vesicles from Broncho-alveolar lavage fluid: a review and comparison of different methods. Respiratory research, 20(1), 240.

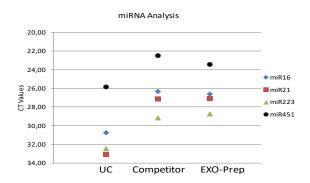




#### EXO-Prep isolates Extracellular Vesicles in one single step from a small volume of sample



18. WB analysis of Alix in EV lysates



19. Profiling of 4 miRNA EV associated.

EXO-Prep is able to isolate EVs form very low volume amount. EVs were isolated from 100  $\mu$ l or 500  $\mu$ l of human plasma, serum and 10 or 20 ml of human urine. 30  $\mu$ g of protein lysates have been used for EV marker analysis (Alix) (fig 18). RNAs were extracted from EVs isolated from 500  $\mu$ l of human plasma and tested for profiling of 4 different miRNA EV associated (fig 19).

